

State-of-the-Art Review

COVID-19 vaccines and coronavirus 19 variants including alpha, delta, and omicron: present status and future directions

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Abstract

Coronavirus disease 2019 (COVID-19) is caused by a coronavirus, also called severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). It has caused urgent global health problems worldwide. The rapid spread of this pathogen, the development of its variants, and the escalating number of patients and their deaths prompted scientists to take up the race for immediate development of COVID-19 vaccines. With regard to the technologies available for vaccine development, mRNA-, inactivated virus-, viral vector-, and protein subunit-based vaccines have been developed thus far. This review article aimed to epitomize the current intelligence and improvements of various vaccines against COVID-19, explicitly those approved for urgent use.

Keywords: COVID-19; Vaccine; SARS-CoV-2; Variant; Alpha; Delta; Omicron.

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1. Introduction

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is a representative of the Coronaviridae family, which constitutes various virulent strains that transmit to humans and animals, comprising SARS-CoV and Middle East respiratory syndrome coronavirus (MERS-CoV).[1] As of 4 March 2022, the coronavirus disease 2019 (COVID-19) pandemic, caused by SARS-CoV-2, has expanded to 220 countries or areas with approximately 440.8 million laboratory-verified cases and 5.98 million deaths (World Health Organization [WHO]);[2] thus, resulting in global social disruption and economic decline. Progress towards availability of secure and efficient vaccines affordable in all countries and areas is essential to put a halt to this pandemic.

A vaccine is essential, particularly, due to the fact that studies have suggested asymptomatic transmission of COVID-19.[3] The expeditious development, circulation, and administration of a vaccine to the population across the world are the most efficient ways to extinguish the pandemic.[4] In general, an effective vaccine takes decades to develop; therefore, rapid availability of effective vaccines for widespread distribution is extraordinary and may yield unpredicted side effects. Much to the public surprise, novel mass-producing platforms, formation-supported virus antigen composition, computational biology, protein manufacturing, and gene integration all have contributed to the advancement of the tools required to produce vaccines with expedition and accuracy.[5]

In order to establish vaccines against SARS-CoV-2, it is of great scientific importance to appropriately interpret the immune systems involved in protection of subjects against the virus.

Mounting evidence has shown the value of both humoral and cellular immunity with regard to the development of a protective effect against COVID-19.[6] For instance, neutralizing antibodies (nAbs) have been reported to reduce the amount of the virus, with a greater response among subjects whose immune system had not yet been infected or who had a high amount of the virus from the beginning.[6] In conjunction with nAbs, T cell responses have an essential role in COVID-19 disease. For example, a study of patients affected by COVID-19 pneumonia showed markedly increased capability of CD4⁺ and CD8⁺ T cells to produce *in vitro* interleukin-17, thereby enhancing the inflammatory response and stimulating neutrophils.[7] Thus, it can be inferred that ideal vaccines should induce both the humoral and cellular immune-mediated reactions in a similar way to the virus that infects the subject in a real world setting.

With respect to global demand for effective vaccines against COVID-19, researchers across the world are in the race of developing COVID-19 vaccines. In accordance with the COVID-19 vaccine tracker and landscape by the WHO, COVID-19 vaccine contenders are classified into 7 categories (Fig. 1), which may be divided into 3 classes: first, protein-based vaccines that use major antigens (i.e., inactivated virus vaccines, virus-like particles [VLPs], and protein subunit vaccines); second, gene-based vaccines that transport genes encoding viral antigens in host cells for *in vivo* manufacturing (i.e., virus-vectored vaccines, DNA vaccines, and mRNA vaccines); and, last, a compound of both protein-based and gene-based access to yield protein immunogens both *in vitro* and *in vivo* (i.e., live-attenuated viral vaccines). As of 4 March, 2022, the WHO detailed 147 vaccines in clinical development including 32 in Phase 3 trials and 10 in Phase 4 trials (WHO; COVID-19 Vaccine tracker). In this review article, we outline and review the targeting spots of utmost scientific value in vaccine contenders with a special focus on the vaccines in use with reported data.

2. Various SARS-CoV-2 molecular proteins as targeting spots for vaccines

SARS-CoV-2 comprises 4 main structural proteins, specifically, 1) spike (S), 2) membrane (M) and 3) envelope (E) proteins; all of them are enclosed in the viral outer surface envelope, and 4) nucleocapsid (N) protein, which is located in the ribonucleoprotein core (Fig. 1).[8] S protein is considered the major protein utilized as a target point in the progress of COVID-19 vaccine development. It is divided into two critical functional subunits; the first one is a membrane-distal S1 subunit composing the globular head of the S protein; and the other one is a membrane-proximal S2 subunit comprising the stalk of the protein. Once engaged with a host cell, the S1 subunit of the virus recognizes and attaches to receptors on the host cell through its receptor-binding domain (RBD); subsequently, the S2 subunit on the virus fuses the envelope of the virus with the host cell membrane, which is a vital process for viral entrance (Fig. 2). In MERS-CoV, SARS-CoV, or SARS-CoV-2, the RBD is positioned in the C-terminal domain of the S1 subunit.[9] In a few coronaviruses, the N-terminal domain (NTD) of the S1 subunit may become utilized for receptor binding. The S2 subunit consists of the fusion peptide (FP), connecting region (CR), heptad repeat 1 (HR1) and HR2 in the vicinity of a central helix as a helix-turn-helix formation. Structural information has reported an image for the realignment of

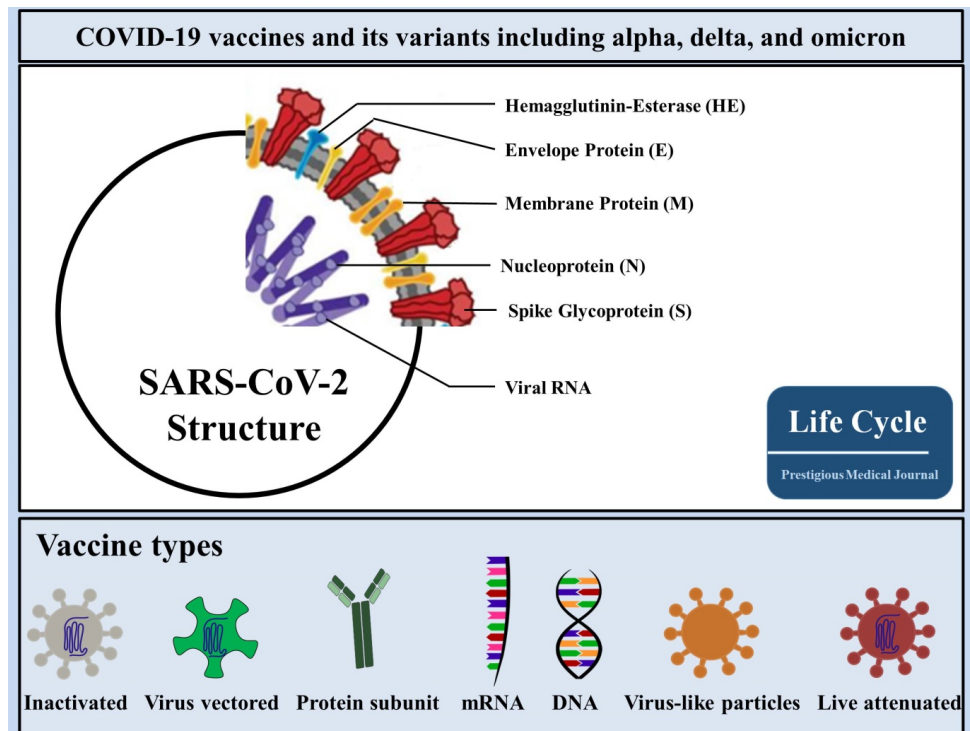


Fig. 1. Various vaccine components of SARS-CoV-2 vaccine contenders.

Inactivated virus vaccines: Viruses are substantially or chemically inactivated; however, conserve the integrity of the virus particle, which plays as the immunogen.

Virus-vectored vaccines: Genes deciphering pathogen antigens are duplicated into non-replicating or replicating virus vectors (such as adenovirus). The antigens are manufactured by transduced host cells after vaccine.

Protein subunit vaccines: This approach constitutes only key viral proteins or peptides that can be synthesized *in vitro* in bacteria, yeast, insect or mammalian cells. The largest number of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) vaccine contenders in both clinical and preclinical stages has been established upon this approach.

DNA and mRNA vaccines: DNA and mRNA vaccines have the convenience of rapid manufacturing against pandemic pathogens. For DNA vaccines, viral antigens ciphered by a recombinant DNA plasmid are manufactured in host cells through a sequential transcription-to-translation mechanism. In contrast, mRNA vaccines are integrated by *in vitro* transcription and they manufacture viral antigen(s) in the cytoplasm via straightforward protein translation *in vivo*.

Virus-like molecule or nano-molecule vaccines: In this design, anatomical viral proteins are expressed alongside each other in order to assemble non-infectious molecules as the vaccine antigen. Although they feature real virions, they are limited in that they lack the virus genome.

Live-attenuated virus vaccines: In this procedure, virus is debilitated by *in vitro* or *in vivo* passage or reverse-genetic mutagenesis. Despite that the resulting virus becomes non-pathogenic or weakly pathogenic, it retains immunogenicity by imitating live virus infection.

SARS-CoV-2 S molecular protein when engaging with the host cell receptor.[10]. The binding of the RBD to its cellular receptor, human angiotensin-converting enzyme 2 (hACE2), induces the disconnection of the S1 subunit and simultaneously institutes the re-bending transformation of the spring-bundled S2 subunit, that bulges the FP at its end for the membrane fusion process. The S2 subunit in its post-fusion configuration convolutes in the formation of a lengthy helical array adjacent to the FP embedded into the host cell membrane. Hypothetically, nAbs can aim at the S protein in order to hinder viral infection at various phases in the course of viral entry. The RBD is the main spot where nAbs hamper viral receptor binding.[11] Further, the S2 subunit is a

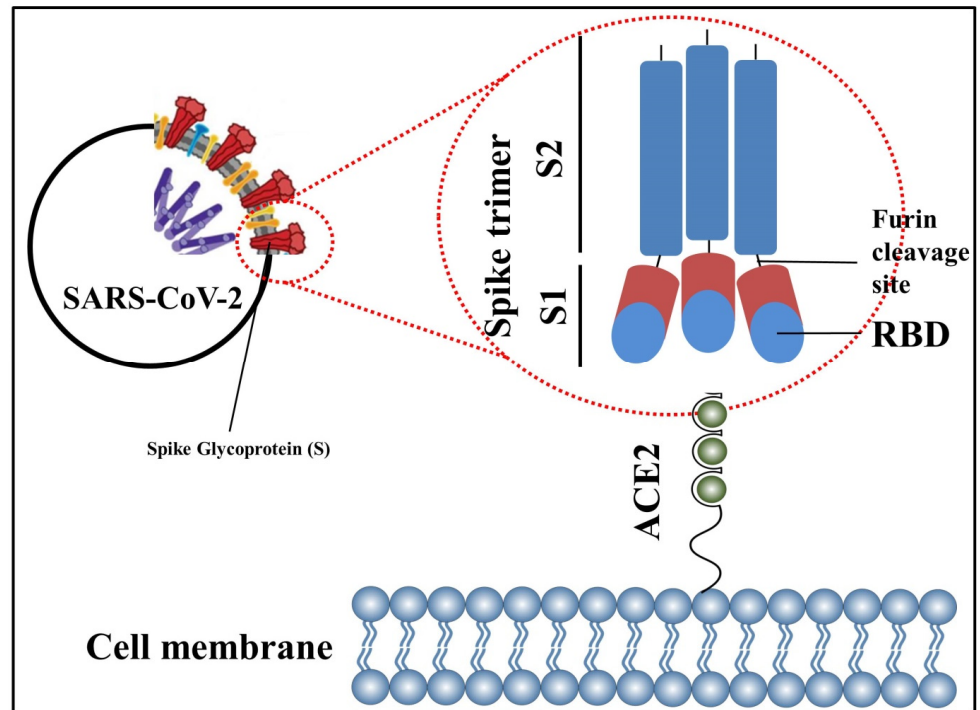


Fig. 2. Major target points being utilized in COVID-19 vaccine contenders.

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) consists of 4 main construction proteins: spike (S), membrane (M) and envelope (E) proteins, that are enclosed on the outer surface of the virus, and nucleocapsid (N) protein, that attaches to viral RNA which is located internal side of the virus. The S protein trimer in its pre-fusion formation is displayed. The S protein constitutes the S1 subunit (that comprises the N-terminal domain [NTD] and the receptor-binding domain [RBD]) (the receptor-binding motif [RBM] inside the RBD is also tagged) and the S2 subunit (that comprises fusion peptide [FP], connecting region [CR], heptad repeat 1 [HR1], heptad repeat [HR2] and central helix [CH]). The SARS-CoV-2 S molecular protein unites its host receptor, the dimeric human angiotensin-converting enzyme 2 (hACE2), through the RBD and detaches the S1 subunits. Breakage at S1–S2 and S2' spots facilitates structural reorganization of the S2 subunit needed for virus–host cell membrane fusion. The S2-trimer in its post-fusion formation is depicted. The RBD is an interesting vaccine targeting point. The formations of an RBD-dimer or RBD-trimer have been reported to augment the immunogenicity of RBD-related vaccines. A balanced S-trimer presented with a C-terminal trimer-tag is a destined spot for vaccines. The pre-fusion S protein is a natural constant status during *in vitro* preparations and likely to convert into its post-fusion formation. Mutation of two residues (K986 and V987) to proline preserves S protein (S-2P) and hinders the pre-fusion to post-fusion systemic structural switch.

possible point for nAbs that impede structural adjustment of the S protein and the embedment of FP essential for virus-host membrane fusion.[12] Furthermore, the S protein is a major spot for eliciting T cell reactions, which is supported by previous studies detailing both CD4⁺ and CD8⁺ T cell epitopes in the S protein of SARS-CoV-2.[13]

3. Virus-vectored and mRNA vaccines for the prevention of SARS-CoV-2

Up to now, three COVID-19 vaccines established upon adenovirus vector with the full-length S protein on the surface have been approved for use; the first one is based on a single-dose of human adenovirus type 5 (Ad5) (i.e., CanSino Biological Inc./Beijing Institute of Biotechnology),[14, 15] the second one uses two-doses of recombinant chimpanzee adenovirus, ChAdOx1-S,(i.e., AstraZeneca/University of Oxford)[16, 17] and the third one is a single-dose

Ad26.COVS vaccine.[18] There are three other vaccines using adenovirus in Phase 3 trials: Recombinant COVID-19 vaccine (adenovirus type 5 vector) for inhalation (Ad5-nCoV-IH),[19] combination of human Ad26 and Ad5 in a prime-boost vaccination regimen (Gam-COVID-Vac Adeno-based [rAd26-S+rAd5-S], Sputnik V),[20] and DeINS1-2019-nCoV-RBD-OPT1 (Intranasal flu-based-RBD). While adenoviral vectors have the advantages of their broad tissue tropism, intrinsic adjuvant elements, and scalability, they also have disadvantages of pre-existing immunity in humans, which may decrease efficiency of the adenoviral vector.[4] To date, there are two DNA vaccine contenders in Phase 3 trial[21] and none in Phase 4 trials.

The S protein is labile when used as a recombinant molecular protein and is likely to reconfigure from its pre-fusion to a post-fusion arrangement, discharging the S1 subunit (Fig. 2). Nonetheless, the S1 subunit is the immunologically predominant immunogen when infected with CoV because of its convenience for immunological perception and its consisting of neutralizing epitopes primarily on its RBD.[13] Methods to preserve the S protein in its pre-fusion arrangement and to augment pre-fusion S protein expression are considered to boost the aspect and capacity of vaccine-induced antibodies pointing at the practically compatible epitopes on the surface of the S1 subunit. Two proline substitutions (2P) at the pinnacle of the central helix and HR1 may hold the S proteins of MERS-CoV, SARS-CoV, and coronavirus HKU1 in the immunologically ideal pre-fusion arrangements (Fig. 2). The ensuing antigen, S-2P, generated higher nAb titers than wild-type S protein in animal models.[22] Based on the earlier encounters with the CoVs, the S-2P arrangement is currently utilized in some vaccine approaches against COVID-19. SARS-CoV-2 S-2P (constituting proline substitutions at residues K986 and V987) is employed as the major immunogen in three gene-based vaccines (i.e., mRNA vaccines by Moderna/National Institute of Allergy and Infectious Diseases [NIAID] administered two-doses with 28 days apart[23] and BioNTech/Pfizer given two-doses 21 days apart[24], and a recombinant Ad26 vaccine manufactured by Janssen Pharmaceutical Companies delivered as a singular dose[18]) (Table 1, Fig. 1). Further, mutations at the separation spots in the S protein are considered to preserve the pre-fusion arrangement of the S protein. S-2P in the Janssen Ad26-vectored vaccine (Ad26.COVS) consists of increased mutations at the S1–S2 polybasic breaking spot from RRAR to SRAG or QQAQ to deliver it protease resistant, which aids to further sustain the S protein in its pre-fusion configuration.[25] All of the aforementioned vaccines, which are in Phase 3 and 4, evoked abundant protective immunity against COVID-19 (Table 1). Of the vaccines mentioned above, the research papers for the two mRNA vaccines (BNT162b2 by BioNTech/Pfizer and mRNA1273 by Moderna/NIAID) and the protein subunit vaccine, NVX-CoV2373[26], inspiringly detailed that these vaccines evoked both great titers of nAbs in recuperating subjects with COVID-19 infection and extensive T cell reactions. Hence, maintaining the S protein in its pre-fusion arrangement displays an efficient route to enhance vaccine competence. Further, DNA or RNA vaccines have distinct advantages that as well as antibody and CD4⁺ T cell responses, they evoke CD8⁺ cytotoxic T cell reactions, which play a critical part for virus eradication.[21] It is of great clinical significance to note that clinical studies by BioNTech/Pfizer and Moderna/NIAID have reported that their mRNA vaccines have approximately 95% effectiveness in the prevention of COVID-19.[23]

Table 1. Current vaccines in use and in progress, number of doses, schedule, route of administration, developer, and their phases

ID	Vaccine platform description	Type of contender vaccine	Number of doses	Schedule	Route of administration	Developers	Phase	References
1	Inactivated virus	CoronaVac; inactivated SARS-CoV-2 vaccine (vero cell)	2	Day 0 + 14	IM	Sinovac Research and Development Co., Ltd	Phase 4	[32]
2	Inactivated virus	Inactivated SARS-CoV-2 vaccine (Vero cell)	2	Day 0 + 21	IM	Sinopharm;	Phase 4	A
3	Inactivated virus	Inactivated SARS-CoV-2 vaccine (Vero cell), vaccine name BBIBP-CorV	2	Day 0 + 21	IM	Sinopharm;	Phase 4	[31]
14	Inactivated virus	SARS-CoV-2 vaccine (vero cells)	2	Day 0 + 28	IM	Institute of Medical Biology + Chinese Academy of Medical Sciences	Phase 3	B
15	Inactivated virus	QazCovid-in® - COVID-19 inactivated vaccine	2	Day 0 + 21	IM	Research Institute for Biological Safety Problems, Rep of Kazakhstan	Phase 3	C
20	Inactivated virus	BBV152 vaccine	2	Day 0 + 14	IM	Bharat Biotech International Limited	Phase 3	D
25	Inactivated virus	Inactivated SARS-CoV-2 vaccine (Vero cell)	2	Day 0 + 28	IM	Shenzhen Kangtai Biological Products Co., Ltd.	Phase 3	E
56	Inactivated virus	VLA2001	2	Day 0 + 21	IM	Valneva, National Institute for Health Research, United Kingdom	Phase 3	F
63	Inactivated virus	TURKOVAC, inactivated virus	2	Day 0 + 21	IM	Erciyes University and the Health Institutes of Turkey (TUSEB)	Phase 3	G
4	Viral vector (non-replicating)	ChAdOx1-S - (AZD1222)	1-2	Day 0 + 28	IM	AstraZeneca + University of Oxford	Phase 4	[15]
5	Viral vector (non-replicating)	Recombinant novel coronavirus vaccine (adenovirus type 5 vector)	1	Day 0	IM	CanSino Biological Inc./Beijing Institute of Biotechnology	Phase 4	[13, 14]
6	Viral vector (non-replicating)	Recombinant COVID-19 vaccine (adenovirus type 5 vector) for Inhalation (Ad5-nCoV-IH)	1	Day 0	IH	CanSino Biological Inc./Beijing Institute of Biotechnology	Phase 3	H

Table 1. Continued

ID	Vaccine platform description	Type of contender vaccine	Number of doses	Schedule	Route of administration	Developers	Phase	References
7	Viral vector (non -replicating)	Gam-COVID-Vac Adeno-based (rAd26-S+rAd5-S)	2	Day 0 + 21	IM	Gamaleya Research Institute ; Health Ministry of the Russian Federation	Phase 3	[10]
8	Viral vector (non -replicating)	Ad26.COVS.2.S	1-2	Day 0 or Day 0 +56	IM	Janssen Pharmaceutical	Phase 4	[17]
38	Viral vector (replicating)	DeINS1-2019-nCoV-RBD-OPT1 (Intranasal flu-based-RBD)	2	Day 0 + 28	IN	University of Hong Kong, Xiamen University and Beijing Wantai Biological Pharmacy	Phase 3	I
9	Protein subunit	SARS-CoV-2 rS/Matrix M1-Adjuvant (Full length recombinant SARS CoV-2)	2	Day 0 + 21	IM	Novavax	Phase 3	[25]
12	Protein subunit	Recombinant SARS-CoV-2 vaccine (CHO cell)	2-3	Day 0 + 28 or	IM	Anhui Zhifei Longcom Biopharmaceutical + Institute of Microbiology, Chinese Academy of Sciences	Phase 3	J
22	Protein subunit	VAT00008: SARS-CoV-2 S protein with adjuvant	2	Day 0 + 21	IM	Sanofi Pasteur + GSK	Phase 3	K
29	Protein subunit	CpG 1018/Alum- adjuvanted recombinant SARS-CoV-2 trimeric S-protein subunit vaccine (SCB-2019)	2	Day 0 + 21	IM	Clover Biopharmaceuticals Inc./Dynavax	Phase 3	L
30	Protein subunit	COVAX-19 [®] Recombinant spike protein + adjuvant	2	Day 0 + 21	IM	Vaxine Pty Ltd./CinnaGen Co.	Phase 3	M
31	Protein subunit	MVC-COV1901 (Spike-2P protein + adjuvant CpG 1018)	2	Day 0 + 28	IM	Medigen Vaccine Biologics + Dynavax + National Institute of Allergy and Infectious Diseases (NIAID)	Phase 4	N
33	Protein subunit	FINLAY-FR-2 anti-SARS-CoV-2 vaccine (RBD chemically conjugated to tetanus toxoid plus adjuvant)	2	Day 0 + 28	IM	Instituto Finlay de Vacunas	Phase 3	O

Table 1. Continued

ID	Vaccine platform description	Type of contender vaccine	Number of doses	Schedule	Route of administration	Developers	Phase	References
34	Protein subunit	EpiVacCorona (EpiVacCorona vaccine based on peptide antigens for the prevention of COVID-19)	2	Day 0 + 21	IM	Federal Budgetary Research Institution State Research Center of Virology and Biotechnology "Vector"	Phase 3	P
35	Protein subunit	RBD (baculovirus production expressed in Sf9 cells) recombinant SARS-CoV-2 vaccine (Sf9 Cell)	2	Day 0 + Day 21 + Day 42	IM	West China Hospital + Sichuan University	Phase 3	Q
55	Protein subunit	CIGB-66 (RBD+aluminium hydroxide)	3	Day 0 + 14 + 28	IM	Center for Genetic Engineering and Biotechnology (CIGB)	Phase 3	R
57	Protein subunit	BECOV2	2	Day 0 + 28	IM	Biological E. Limited	Phase 3	S
60	Protein subunit	Recombinant Sars-CoV-2 spike protein, aluminum adjuvanted (Nanocovax)	2	Day 0 + 21	IM	Nanogen Pharmaceutical Biotechnology	Phase 3	T
61	Protein subunit	Recombinant protein vaccine S-268019 (using Baculovirus expression vector system)	2	Day 0 + 21	IM	Shionogi	Phase 3	U
65	Protein subunit	GBP510, a recombinant surface protein vaccine with adjuvant AS03 (aluminium hydroxide)	2	Day 0 + 28	IM	SK Bioscience Co., Ltd. and CEPI	Phase 3	V
66	Protein subunit	Razi Cov Pars, recombinant spike protein	3	Day 0 + 21 + 51	IM and IN	Razi Vaccine and Serum Research Institute	Phase 3	W
91	Protein subunit	Recombinant SARS-CoV-2 fusion protein vaccine (V-01)	2	Day 0 + 21	IM	Livzon Pharmaceutical	Phase 3	X
106	Protein subunit	RBD protein recombinant SARS-CoV-2 vaccine (Noora vaccine)	3	Day 0 + 21 + 35	IM	Bagheiat-allah University of Medical Sciences/AmitisGen	Phase 3	Y
10	RNA based vaccine	mRNA-1273	2	Day 0 + 28	IM	Moderna + National Institute of Allergy and Infectious Diseases (NIAID)	Phase 4	[22]
11	RNA based vaccine	BNT162b2 (3 LNP-mRNAs), also known as "Comirnaty"	2	Day 0 + 21	IM	Pfizer/BioNTech + Fosun Pharma	Phase 4	[23]

Table 1. Continued

ID	Vaccine platform description	Type of contender vaccine	Number of doses	Schedule	Route of administration	Developers	Phase	References
13	RNA based vaccine	CVnCoV vaccine	2	Day 0 + 28	IM	CureVac AG	Phase 3	Z
40	RNA based vaccine	SARS-CoV-2 mRNA vaccine (ARCoV)	2	Day 0 + 14 or Day 0 + 28	IM	Academy of Military Science (AMS), Walvax Biotechnology and Suzhou Abogen Biosciences	Phase 3	AA
78	RNA based vaccine	mRNA-1273.351.	3	Day 0 or Day 0 + 28	IM	Moderna + National Institute of Allergy and Infectious Diseases (NIAID)	Phase 4	BB
115	RNA based vaccine	ARCT-154 mRNA vaccine	2	Day 0 + 28	IM	Arcturus Therapeutics, Inc.	Phase 3	CC
16	DNA based vaccine	INO-4800+electroporation	2	Day 0 + 28	ID	Inovio Pharmaceuticals + International Vaccine Institute + Advaccine (Suzhou) Biopharmaceutical Co., Ltd	Phase 3	[20]
18	DNA based vaccine	nCov vaccine	3	Day 0 + 28 + 56	ID	Zyodus Cadila	Phase 3	DD
41	Virus like particle	Coronavirus-Like Particle COVID-19 (CoVLP)	2	Day 0 + 21	IM	Medicago Inc.	Phase 3	EE
53	Live attenuated virus	COVI-VAC	1-2	Day 0 or Day 0 + 28	IN	Codagenix/Serum Institute of India	Phase 3	CC

APC , antigen presenting cell; ID, intradermal; IM, intramuscular; IN, intranasal; IV, inactivated virus; LAV, live-attenuated virus; PS, protein subunit; VVnr, viral vector (non-replicating); VLP, virus like particle; VVr, viral vector (replicating).

A. <http://www.chictr.org.cn/showprojen.aspx?proj=56651>

B. <http://www.chictr.org.cn/showprojen.aspx?proj=56651>

C. <https://clinicaltrials.gov/ct2/show/NCT04691908>

D. <https://clinicaltrials.gov/ct2/show/NCT04641481>

E. <https://clinicaltrials.gov/ct2/show/NCT04852705>

F. <https://clinicaltrials.gov/ct2/show/NCT04864561>

G. <https://www.clinicaltrials.gov/ct2/show/NCT04942405?term=TURKOVAC&draw=1>.

H. <https://clinicaltrials.gov/ct2/show/NCT05124561>

I. Chinese Clinical Trial Register (ChiCTR) - The world health organization international clinical trials registered organization registered platform

J. <https://www.chictr.org.cn/showprojen.aspx?proj=133228>.

K. [https://pactr.samrc.ac.za/TrialDisplay.aspx?TrialID=13475 2020](https://pactr.samrc.ac.za/TrialDisplay.aspx?TrialID=13475%2020)

L. [https://clinicaltrials.gov/ct2/show/NCT05012787 2021](https://clinicaltrials.gov/ct2/show/NCT05012787%2021)

M. [https://en.irct.ir/trial/57559 2021](https://en.irct.ir/trial/57559%2021)

N. [https://clinicaltrials.gov/ct2/show/NCT05079633 2021](https://clinicaltrials.gov/ct2/show/NCT05079633%2021)

O. Eugenia Toledo-Romani M, Garcia-Carmenate M, Silva V, Baldoquin-Rodriguez W, Martínez Pérez M, Rodríguez Gonzalez M, et al. Efficacy and safety of SOBERANA 02, a COVID-19 conjugate vaccine in heterologous three-dose combination. medRxiv. 2021.

P. <https://www.clinicaltrials.gov/ct2/show/NCT04780035?term=vaccine&cond=Covid19&draw=2>

Q. [https://clinicaltrials.gov/ct2/show/NCT04904471 2021](https://clinicaltrials.gov/ct2/show/NCT04904471%2021)

R. [https://rpcec.sld.cu/trials/RPCEC00000359-En 2021](https://rpcec.sld.cu/trials/RPCEC00000359-En%2021)

S. [http://www.ctri.nic.in/Clinicaltrials/pmaindet2.php?trialid=59772 2022](http://www.ctri.nic.in/Clinicaltrials/pmaindet2.php?trialid=59772%2022)

T. [https://clinicaltrials.gov/ct2/show/NCT04922788 2021](https://clinicaltrials.gov/ct2/show/NCT04922788%2021)

U. <https://clinicaltrials.gov/ct2/show/NCT05212948>
V <https://clinicaltrials.gov/ct2/show/NCT05007951> 2021
W. <https://en.ircct.ir/trial/57980>
X. <https://clinicaltrials.gov/ct2/show/NCT05096832>
Y. <https://en.ircct.ir/trial/60796>.
Z. <https://clinicaltrials.gov/ct2/show/NCT04674189>
AA. <https://clinicaltrials.gov/ct2/show/NCT04847102>
BB. <https://www.clinicaltrialsregister.eu/ctr-search/trial/2021-000930-32/BE>
CC. <https://www.isrctn.com/ISRCTN15779782>
DD. <http://ctri.nic.in/Clinicaltrials/showallp.php?mid1=45306&EncHid=&userName=Zyodus>
EE. <https://clinicaltrials.gov/ct2/show/NCT04636697>

The RBD attaches to the host receptor through a receptor-binding motif (RBM) on its external subdomain in SARS-CoV-2.[9] Since glycans cover the surface of the S protein, with the exception of the RBD, and shield it from antibodies, most SARS-CoV-2 nAbs attach to the RBD and prevent the RBD–hACE2 combination, thereby reducing viral adherence.[27] In this regard, there are a handful of clinical trials using RBD-targeted vaccines in Phase 3 (Fig. 1).[28]

In addition, the S1-NTD consists of epitopes for CoV nAbs detected in recovering subjects with COVID-19 and has been recognized as a possible major targeting point in CoV vaccines.[29] NTD-pointing nAbs typically do not explicitly hinder receptor binding; instead, they hamper receptor binding or constrain the S protein structural shifts necessary for the pre-fusion to post-fusion conversion. However, SARS-CoV-2 NTD-binding nAbs typically show less neutralizing potency compared to RBD-specific nAbs.[12] In this regard, NTD-targeting vaccines against COVID-19 have not been developed.

With regard to the S2 subunit, although there is some evidence reporting the S2 subunit as a targeting point for COVID-19 vaccine, S2 subunit-targeting antibodies retrieved from recovering patients reported less neutralizing activities against SARS-CoV-2 than RBD-targeting antibodies.[30]

CoV M and E proteins are hardly able to produce humoral responses, likely as a result of their small molecular sizes and limited ectodomains, which could be recognized by immune cells.[12] (Fig. 2) Although there is some scientific evidence that the sequence identity of M or E proteins in SARS-CoV-2 is considerably higher than for the S protein and RBD, proposing the potential of M and E proteins as targeting points for cross-reactive T cells,[12] there have been no solid studies aiming to develop vaccines using the M or E proteins. The N protein is the most abundant viral protein and is able to produce an immune response following CoV infections.[31] Hence, it is a main target protein not only for evoking antibody reactions, but also for T cell responses.[31] Despite that some scientific evidence shows that N-specific antibodies were found to defend mice against mouse hepatitis virus, a mouse CoV, via Fc-mediated effector functions, anti-N immune responses were not able to show protection against SARS-CoV-2 infection in animal models.[31] Taken together, there is one vaccine using a protein subunit in Phase 4 (i.e., MVC-COV1901 [Spike-2P protein + adjuvant CpG 1018]) and 17 vaccines in Phase 3.[26]

Inactivated virus and live-attenuated virus vaccines utilize the whole virus as a vaccine target. They encompass complete structural proteins (i.e., S, M, E, and N proteins) and vaccines using live-attenuated virus may develop non-architectural and component proteins *in vivo*.

Hence, these vaccine contenders are able to evoke greater humoral and T cell responses than the aforementioned ones, that are established upon a singular protein or protein particles. There are three vaccines using inactivated virus in Phases 4; first, inactivated SARS-CoV-2 vaccine (Vero cell), vaccine named BBIBP-CorV,[32] second, CoronaVac; inactivated SARS-CoV-2 vaccine (Vero cell),[33] and third, inactivated SARS-CoV-2 vaccine (Vero cell) and six others in Phase 3.[34] Since these vaccine formulations are inadequate for replication and safer than live-attenuated vaccines, their inactivation leads to decreased immunogenicity and the need for multiple-dose regimens in order to institute resilient immunity. In addition, they frequently need adjuvants to vaccinate the aging population by reason of immune senescence.[35] In contrast to inactivated virus, live-attenuated vaccines (LAV) are reproducing, but avirulent viruses. LAV administration requires a single-dose to generate immunity. LAVs have the risk of transferring the virus and/or conversion to a pathogenic variety, reactivation in immune-compromised subjects or recombination with alike-viruses disseminating in the general population.[5] Thus far, there has been one SARS-CoV-2 live-attenuated vaccine in Phase 3.[35]

4. Conclusions

To date, several vaccines for SARS-CoV-2 and its variants including alpha, delta, and omicron are available with safety and scientifically reasonable levels of protection efficacy for COVID-19 (i.e., BioNTech/Pfizer and Moderna/NIAID), although more needs to be studied with regard to the effect of mRNA vaccines on the omicron variant. Of note, it is challenging, in a clinical setting, to accurately analyze the effectiveness and the duration of protection by the various vaccines due to the fact that there are no standardized methods to measure neutralization levels. In addition, researchers and the world should find an expeditious way to discover novel COVID-19 vaccines which will be effective for the newly discovered omicron variant and possibly more variants in the future in order to keep this disastrous virus under control.

Capsule Summary

This review article aimed to epitomize the current intelligence and improvements of various vaccines against COVID-19, explicitly those approved for urgent use.

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Conflicts of Interest

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References

1. Lee SW, Yuh WT, Yang JM, Cho YS, Yoo IK, Koh HY, et al. Nationwide results of COVID-19 contact tracing in South Korea: individual participant data from an epidemiological survey. *JMIR Medical Informatics*. 2020;8(8):e20992.
2. Kim SY. Nationwide COVID-19 vaccination coverage and COVID-19 incidence in South Korea, January 2022: A National Official Report. *Life Cycle*. 2022;2:e2.
3. Sutton D, Fuchs K, D'Alton M, Goffman D. Universal Screening for SARS-CoV-2 in Women Admitted for Delivery. *The New England Journal of Medicine*. 2020;382(22):2163-4.
4. Shin MD, Shukla S, Chung YH, Beiss V, Chan SK, Ortega-Rivera OA, et al. COVID-19 vaccine development and a potential nanomaterial path forward. *Nature Nanotechnology*. 2020;15(8):646-55.
5. Dai L, Gao GF. Viral targets for vaccines against COVID-19. *Nature Reviews Immunology*. 2021;21(2):73-82.
6. Weinreich DM, Sivapalasingam S, Norton T, Ali S, Gao H, Bhore R, et al. REGN-COV2, a neutralizing antibody cocktail, in outpatients with Covid-19. *The New England Journal of Medicine*. 2021;384(3):238-51.
7. De Biasi S, Meschiari M, Gibellini L, Bellinazzi C, Borella R, Fidanza L, et al. Marked T cell activation, senescence, exhaustion and skewing towards TH17 in patients with COVID-19 pneumonia. *Nature Communications*. 2020;11(1):3434.
8. Srinivasan S, Cui H, Gao Z, Liu M, Lu S, Mkandawire W, et al. Structural genomics of SARS-CoV-2 indicates evolutionary conserved functional regions of viral proteins. *Viruses*. 2020;12(4).
9. Yuan Y, Cao D, Zhang Y, Ma J, Qi J, Wang Q, et al. Cryo-EM structures of MERS-CoV and SARS-CoV spike glycoproteins reveal the dynamic receptor binding domains. *Nature Communications*. 2017;8:15092.
10. Cai Y, Zhang J, Xiao T, Peng H, Sterling SM, Walsh RM, Jr., et al. Distinct conformational states of SARS-CoV-2 spike protein. *Science (New York, NY)*. 2020;369(6511):1586-92.
11. Premkumar L, Segovia-Chumbez B, Jadi R, Martinez DR, Raut R, Markmann A, et al. The receptor binding domain of the viral spike protein is an immunodominant and highly specific target of antibodies in SARS-CoV-2 patients. *Science Immunology*. 2020;5(48).
12. Chi X, Yan R, Zhang J, Zhang G, Zhang Y, Hao M, et al. A neutralizing human antibody binds to the N-terminal domain of the spike protein of SARS-CoV-2. *Science (New York, NY)*. 2020;369(6504):650-5.
13. Liu WJ, Zhao M, Liu K, Xu K, Wong G, Tan W, et al. T-cell immunity of SARS-CoV: Implications for vaccine development against MERS-CoV. *Antiviral Research*. 2017;137:82-92.

14. Zhu FC, Guan XH, Li YH, Huang JY, Jiang T, Hou LH, et al. Immunogenicity and safety of a recombinant adenovirus type-5-vectored COVID-19 vaccine in healthy adults aged 18 years or older: a randomised, double-blind, placebo-controlled, phase 2 trial. *Lancet* (London, England). 2020;396(10249):479-88.
15. Zhu FC, Li YH, Guan XH, Hou LH, Wang WJ, Li JX, et al. Safety, tolerability, and immunogenicity of a recombinant adenovirus type-5 vectored COVID-19 vaccine: a dose-escalation, open-label, non-randomised, first-in-human trial. *Lancet* (London, England). 2020;395(10240):1845-54.
16. Folegatti PM, Ewer KJ, Aley PK, Angus B, Becker S, Belij-Rammerstorfer S, et al. Safety and immunogenicity of the ChAdOx1 nCoV-19 vaccine against SARS-CoV-2: a preliminary report of a phase 1/2, single-blind, randomised controlled trial. *Lancet* (London, England). 2020;396(10249):467-78.
17. Ramasamy MN, Minassian AM, Ewer KJ, Flaxman AL, Folegatti PM, Owens DR, et al. Safety and immunogenicity of ChAdOx1 nCoV-19 vaccine administered in a prime-boost regimen in young and old adults (COV002): a single-blind, randomised, controlled, phase 2/3 trial. *Lancet* (London, England). 2021;396(10267):1979-93.
18. Sadoff J, Gray G, Vandebosch A, Cárdenas V, Shukarev G, Grinsztejn B, et al. Safety and efficacy of single-dose Ad26.COV2.S vaccine against Covid-19. *The New England Journal of Medicine*. 2021;384(23):2187-201.
19. Wu S, Huang J, Zhang Z, Wu J, Zhang J, Hu H, et al. Safety, tolerability, and immunogenicity of an aerosolised adenovirus type-5 vector-based COVID-19 vaccine (Ad5-nCoV) in adults: preliminary report of an open-label and randomised phase 1 clinical trial. *The Lancet Infectious Diseases*. 2021;21(12):1654-64.
20. Logunov DY, Dolzhikova IV, Shcheblyakov DV, Tukhvatulin AI, Zubkova OV, Dzharullaeva AS, et al. Safety and efficacy of an rAd26 and rAd5 vector-based heterologous prime-boost COVID-19 vaccine: an interim analysis of a randomised controlled phase 3 trial in Russia. *Lancet* (London, England). 2021;397(10275):671-81.
21. Smith TRF, Patel A, Ramos S, Elwood D, Zhu X, Yan J, et al. Immunogenicity of a DNA vaccine candidate for COVID-19. *Nature Communications*. 2020;11(1):2601.
22. Lien CE, Lin YJ, Chen C, Lian WC, Kuo TY, Campbell JD, et al. CpG-adjuvanted stable prefusion SARS-CoV-2 spike protein protected hamsters from SARS-CoV-2 challenge. *Scientific Reports*. 2021;11(1):8761.
23. Baden LR, El Sahly HM, Essink B, Kotloff K, Frey S, Novak R, et al. Efficacy and safety of the mRNA-1273 SARS-CoV-2 vaccine. *The New England Journal of Medicine*. 2021;384(5):403-16.
24. Mahase E. Covid-19: Pfizer vaccine efficacy was 52% after first dose and 95% after second dose, paper shows. *BMJ* (Clinical research ed). 2020;371:m4826.
25. Bangaru S, Ozorowski G, Turner HL, Antanasijevic A, Huang D, Wang X, et al. Structural analysis of full-length SARS-CoV-2 spike protein from an advanced vaccine candidate. *Science* (New York, NY). 2020;370(6520):1089-94.
26. Toback S, Galiza E, Cosgrove C, Galloway J, Goodman AL, Swift PA, et al. Safety, immunogenicity, and efficacy of a COVID-19 vaccine (NVX-CoV2373) co-administered with seasonal influenza vaccines: an exploratory substudy of a randomised, observer-blinded, placebo-controlled, phase 3 trial. *The Lancet Respiratory Medicine*. 2022;10(2):167-

- 79.
27. Liu Z, Xu W, Xia S, Gu C, Wang X, Wang Q, et al. RBD-Fc-based COVID-19 vaccine candidate induces highly potent SARS-CoV-2 neutralizing antibody response. *Signal Transduction and Targeted Therapy*. 2020;5(1):282.
 28. Nayak SK. Inhibition of S-protein RBD and hACE2 interaction for control of SARSCoV-2 infection (COVID-19). *Mini Reviews in Medicinal Chemistry*. 2021;21(6):689-703.
 29. Liu L, Wang P, Nair MS, Yu J, Rapp M, Wang Q, et al. Potent neutralizing antibodies against multiple epitopes on SARS-CoV-2 spike. *Nature*. 2020;584(7821):450-6.
 30. Wec AZ, Wrapp D, Herbert AS, Maurer DP, Haslwanter D, Sakharkar M, et al. Broad neutralization of SARS-related viruses by human monoclonal antibodies. *Science (New York, NY)*. 2020;369(6504):731-6.
 31. Sariol A, Perlman S. Lessons for COVID-19 Immunity from other coronavirus infections. *Immunity*. 2020;53(2):248-63.
 32. Xia S, Zhang Y, Wang Y, Wang H, Yang Y, Gao GF, et al. Safety and immunogenicity of an inactivated SARS-CoV-2 vaccine, BBIBP-CorV: a randomised, double-blind, placebo-controlled, phase 1/2 trial. *The Lancet Infectious Diseases*. 2021;21(1):39-51.
 33. Jara A, Undurraga EA, González C, Paredes F, Fontecilla T, Jara G, et al. Effectiveness of an inactivated SARS-CoV-2 vaccine in Chile. *The New England Journal of Medicine*. 2021;385(10):875-84.
 34. Ella R, Reddy S, Blackwelder W, Potdar V, Yadav P, Sarangi V, et al. Efficacy, safety, and lot-to-lot immunogenicity of an inactivated SARS-CoV-2 vaccine (BBV152): interim results of a randomised, double-blind, controlled, phase 3 trial. *Lancet (London, England)*. 2021;398(10317):2173-84.
 35. Ciabattini A, Nardini C, Santoro F, Garagnani P, Franceschi C, Medaglini D. Vaccination in the elderly: the challenge of immune changes with aging. *Seminars in Immunology*. 2018;40:83-94.